

Unsupervised Deep Learning for Microcopy Imaging Restoration

Nianyi Li

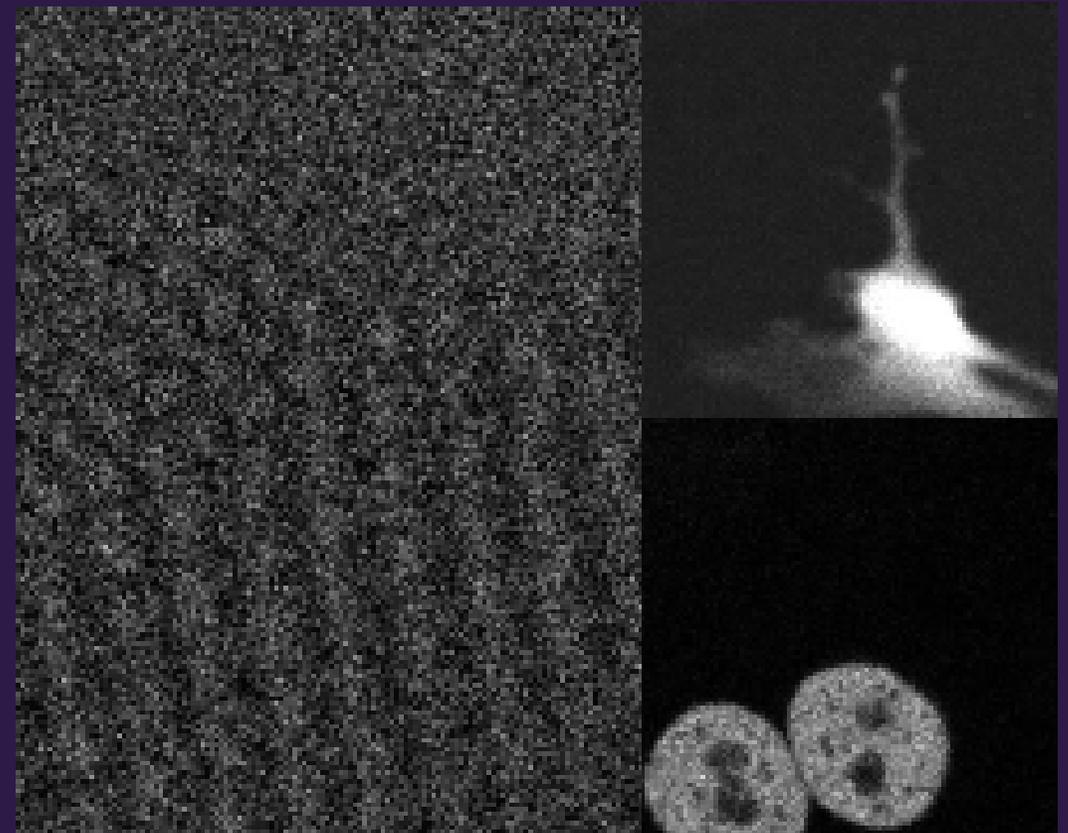
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Motivation

Challenges with Microscopy Imaging

- Low signal-to-noise ratio
- Different noise types
- Multi-modality
- Motion Blur

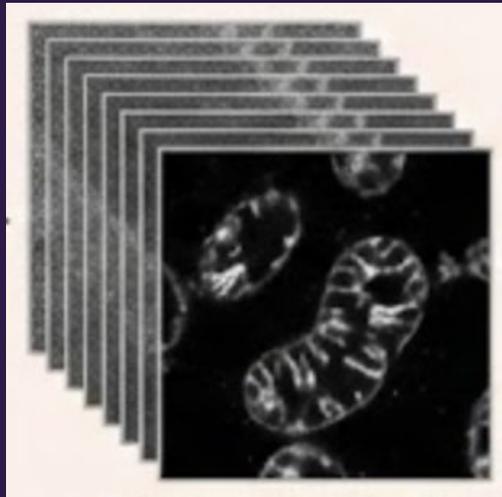


Motivation

Challenges with Microscopy Imaging

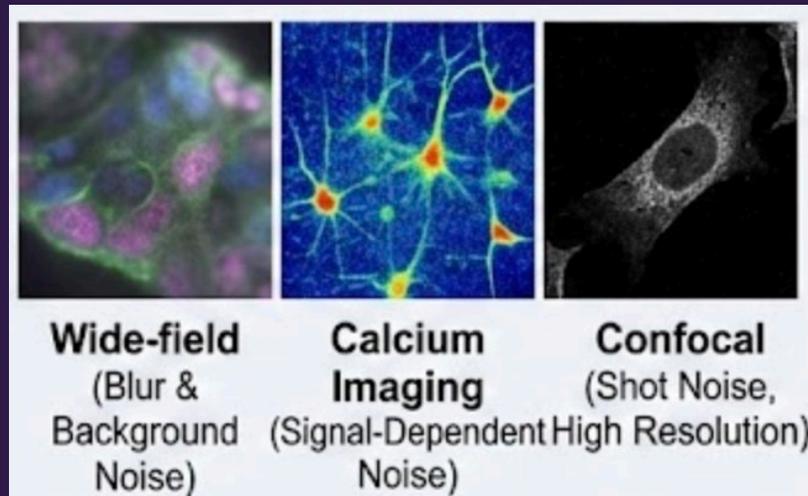
Small data problem:

Most denoising models require thousands of frames



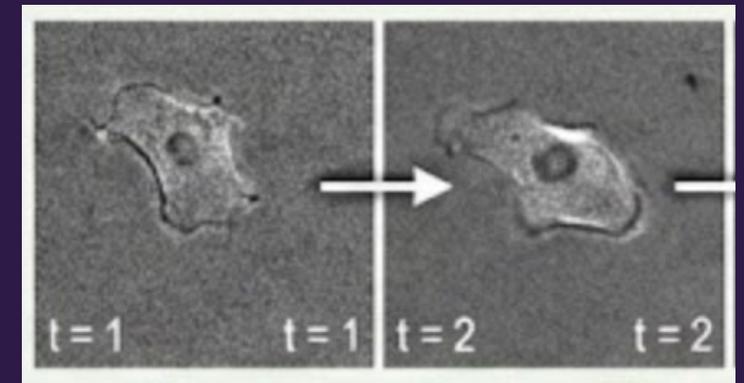
Modality diversity:

Noise varies across wide-field, calcium, and confocal imaging, limiting cross-modality generalization.



Temporal complexity:

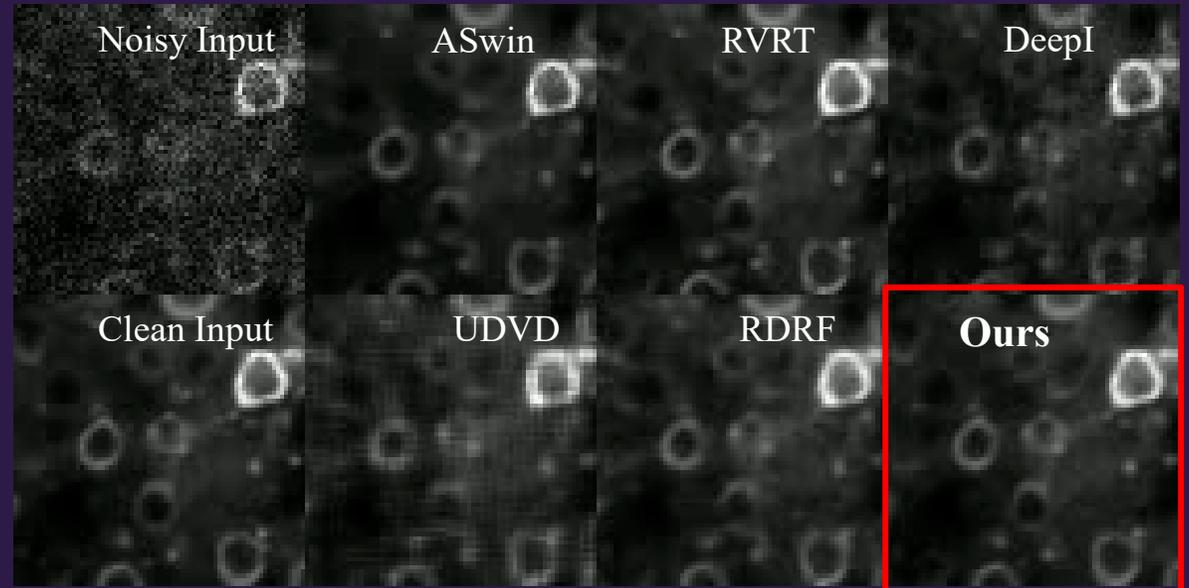
Balancing noise removal and motion preservation in complex or rapid motion scenarios.



Moving cells

Related Works

- Employs Blindspot mechanism to prevent identity noise mapping
- Extensively exploits neighboring pixels, which can lead to overfitting in microscopy images [1,2,3].
- Most existing denoising techniques are primarily designed for Gaussian noise [4,5,6].



[1] Lecoq et al., Removing independent noise in systems neuroscience data using deepinterpolation. Nature methods, 2021.

[2] Sheth et al., Unsupervised deep video denoising. ICCV, 2022

[3] Wang et al., Recurrent self-supervised video denoising with denser receptive field. In Proceedings of the 31st ACM ICM, 2023.

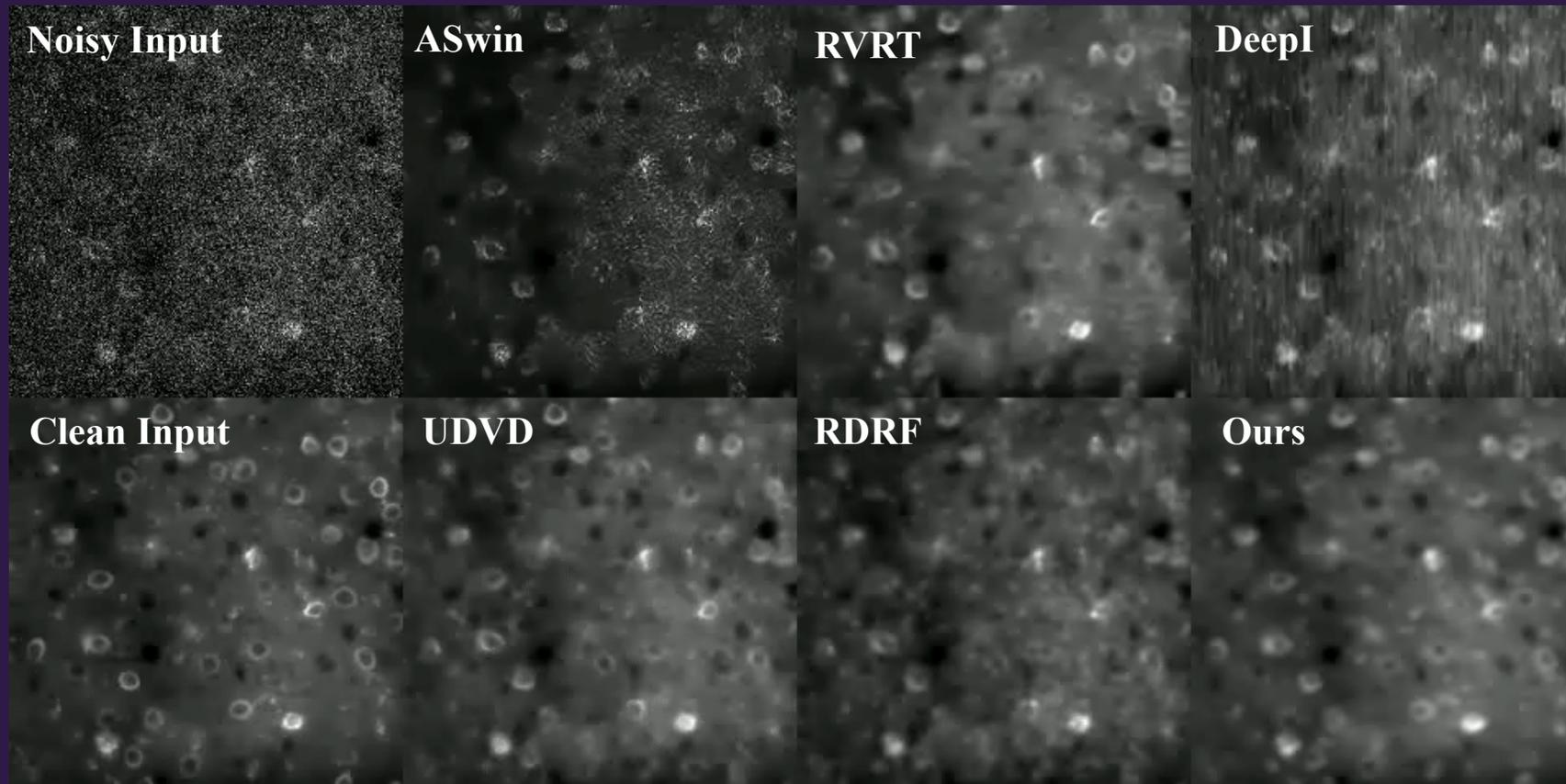
[4] Song et al., Neural anatomy and optical microscopy (naomi) simulation for evaluating calcium imaging methods. Journal of neuroscience methods 2

[5] Liang et al., Recurrent video restoration transformer with guided deformable attention. Neurips 2022

[6] Lindner et al., Lightweight video denoising using aggregated shifted window attention. ICCV, 2023.

Previous Solutions

Unsupervised/Self-Supervised Deep Learning solutions are not efficient and accurate



Outline

- Unsupervised Microscopy Video Denoising
- Unsupervised Denoising for Multi-Modal Microscopy Imaging for Small Data Scenarios

Unsupervised Microscopy Video Denoising

Our solution – Removing the Noise



- **Calcium imaging** is used for recording neural activity over a period
- Presence of **noise can impact the detection** of potential **neural activities** in the recordings
- Some **learning-based approaches** [1,2] for denoising **require** noisy/clean pair of images to train. However, **ground truth** clean images are usually unavailable in medical imaging
- We used an **unsupervised** learning-based approach trained on noisy data by learning the spatio-temporal relationship in the data

[1] HC. Burger, CJ. Schuler, and S. Harmeling (2012). Image denoising: Can plain neural networks compete with bm3d? CVPR

[2] Pascal V., Hugo L., Isabelle L., Y., and Pierre-Antoine M. (2010). Stacked denoising autoencoders: Learning useful representations in a deep network with a local denoising criterion. JMLR

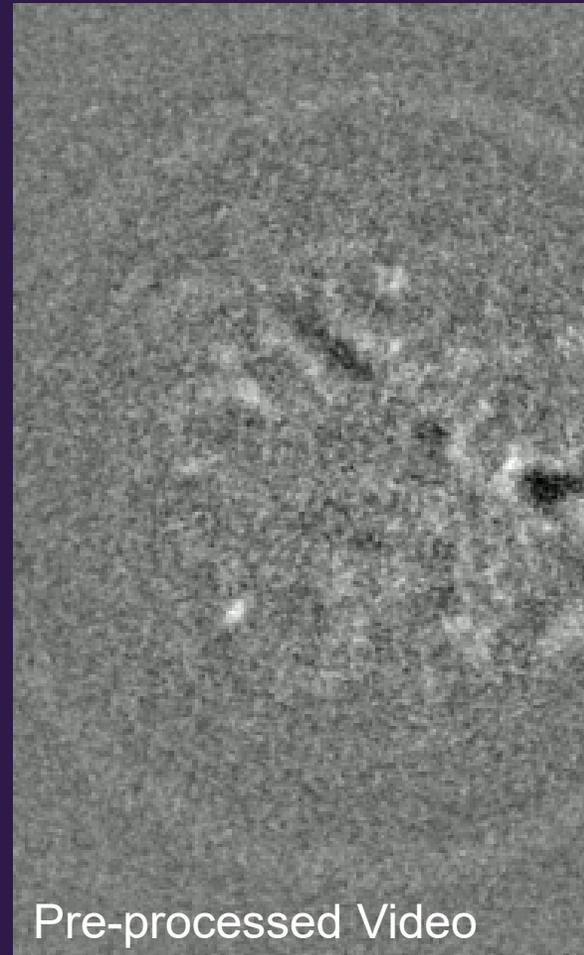
Pre-processed Denoised Frame



However, it will remove the subtle neuron signals and blur out the response area.

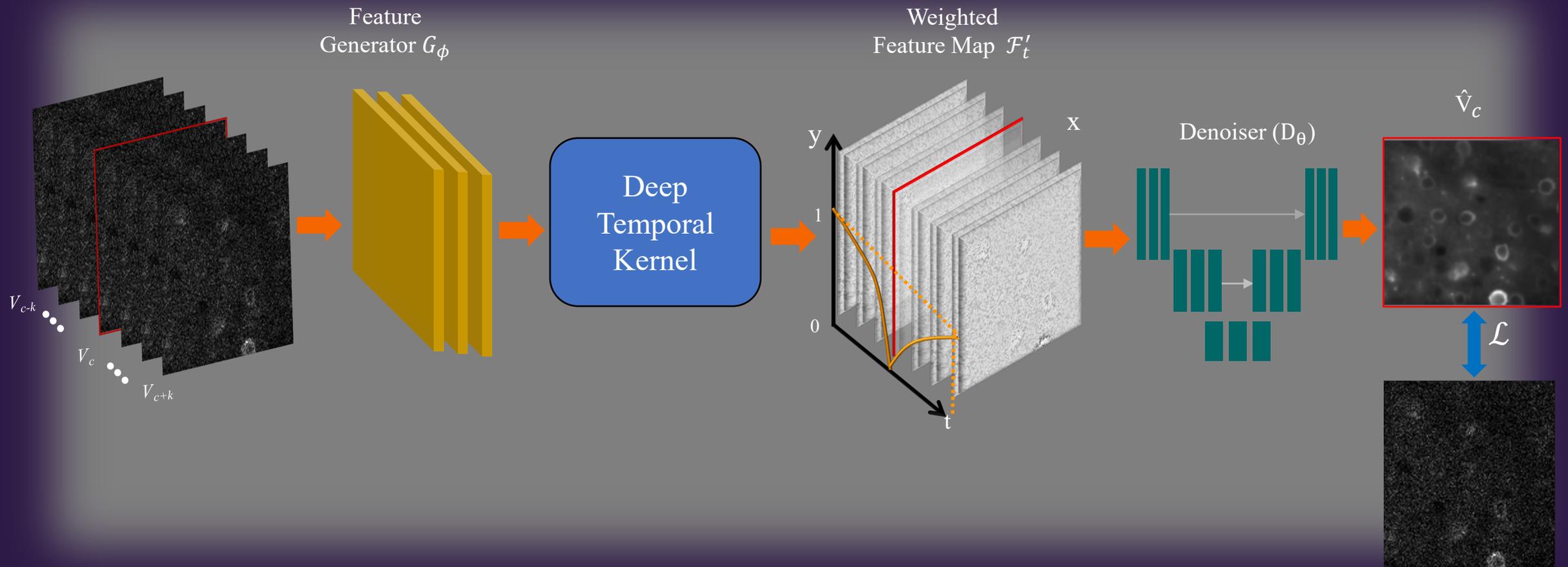
Our solution: Pre-process the input video to amplify the neuron response, then apply denoise algorithm

Pre-processed Denoised Frame



Our solution: Pre-process the input video to amplify the neuron response, then apply denoise algorithm

Our Method



Method

- **DeepTemporal Filter (ω)**

- Linear signal filter (ω) to adjust feature map weights based on temporal proximity to central frame

$$\omega(\cdot) = \left[\left\{ \frac{k-i}{k} \right\}_{i=0}^{k-1}, \left\{ \frac{i}{k} \right\}_{i=1}^k \right]$$

$$\omega(\cdot) = \{1, \dots, 0, \dots, 1\}$$

where

- $k = \lfloor N/2 \rfloor$ and N is the number of frames in the input
- $\{\omega_t | t = 1, \dots, N\}$

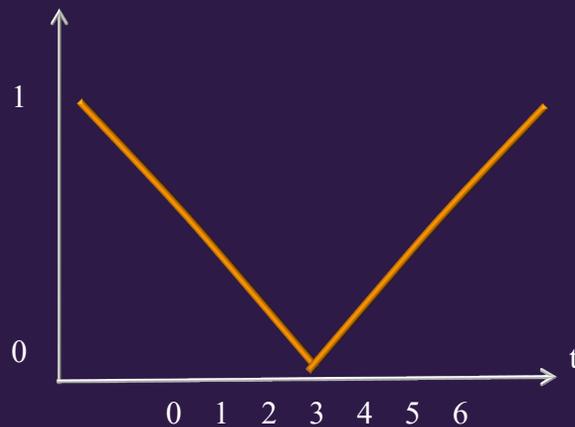


Fig 1. DeepTemporal filter shape

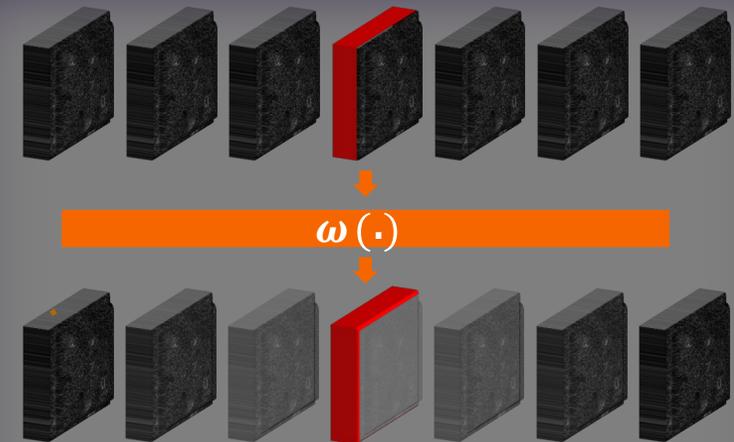


Fig 2. Illustration of our DeepTemporal filter

Method

- **Feature Generator**

- Given a stack consecutive N input frame $\mathcal{V}_t \in \mathbb{R}^{N \times H \times W \times 1}$
- Applies group convolution layers to extract features of each frame.

$$\mathcal{F}_t = G_\phi(\mathcal{V}_t) \quad \mathcal{F}_t \in \mathbb{R}^{N \times H \times W \times C}$$

- **DeepTemporal Filter**

$$\mathcal{F}'_t = \omega_t \odot \mathcal{F}_t \quad \forall t \in \{1, \dots, N\} \quad \mathcal{L} = \left\| \hat{\mathcal{V}}_c - \mathcal{V}_c \right\|_2$$
$$\{\mathcal{F}'_t | t = 1, \dots, c, \dots, N\}$$

- **Denoiser**

- UNet architecture. Comprises of 3 encoder layers and 2 decoder layers

$$\hat{\mathcal{V}}_c = \mathcal{D}_\theta([\mathcal{F}'_1, \dots, \mathcal{F}'_N])$$

Experiment

Dataset

Simulated two-photon Calcium Imaging [1]

One-photon Calcium Imaging [2]

Fluorescence Microscopy [3]

Evaluation Metrics

PSNR

SSIM

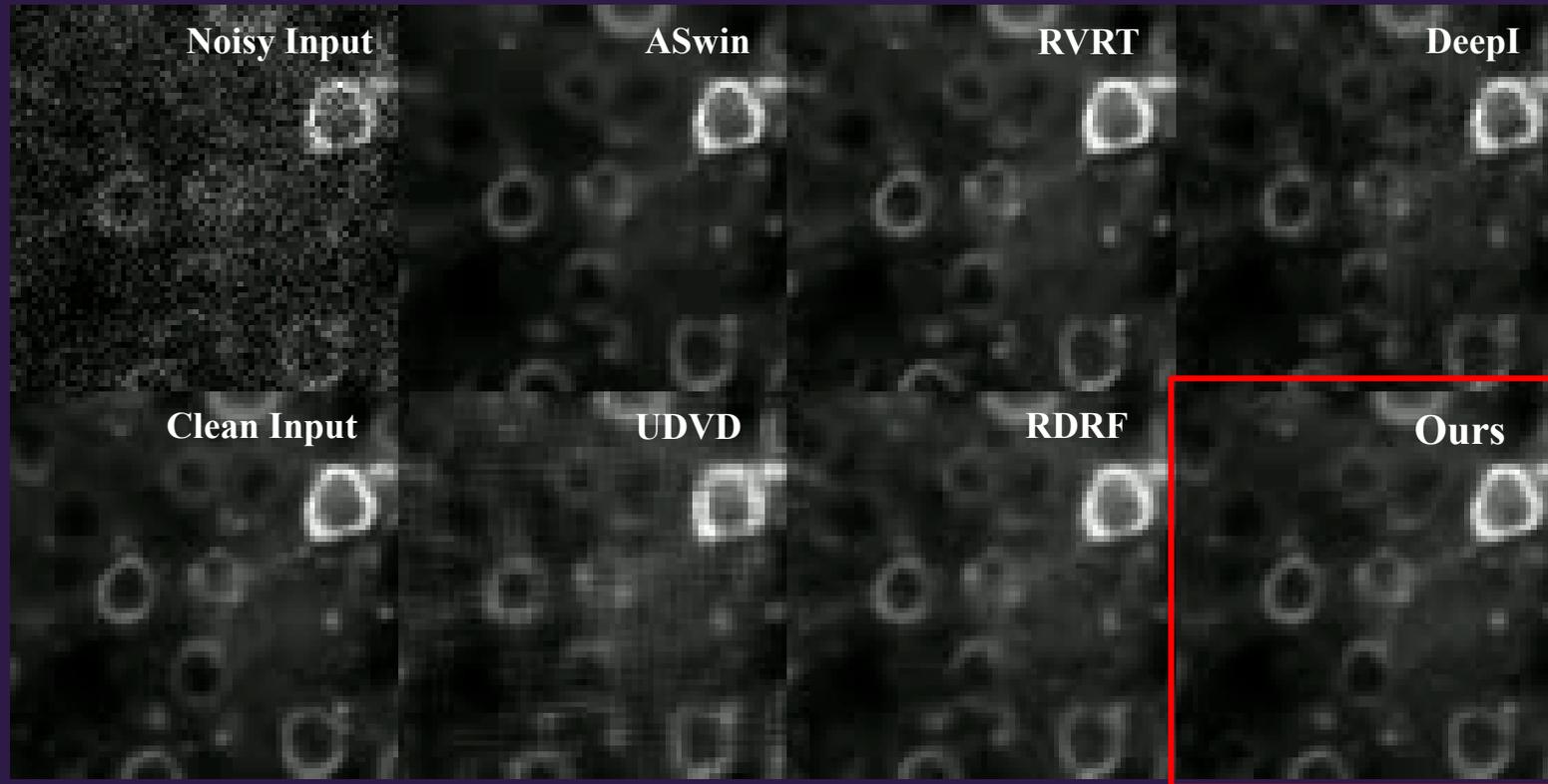
[1] Song et al., Neural anatomy and optical microscopy (naomi) simulation for evaluating calcium imaging methods. Journal of neuroscience methods 2021

[2] Ulman et al., An objective comparison of cell-tracking algorithms. Nature methods 2017.

[3] Kalivas Laboratory, The Medical University of South Carolina.

Results

Simulated two-photon calcium imaging



[1] Lecoq et al., Removing independent noise in systems neuroscience data using deepinterpolation. Nature methods, 2021.

[2] Sheth et al., Unsupervised deep video denoising. ICCV, 2022

[3] Wang et al., Recurrent self-supervised video denoising with denser receptive field. In Proceedings of the 31st ACM ICM, 2023.

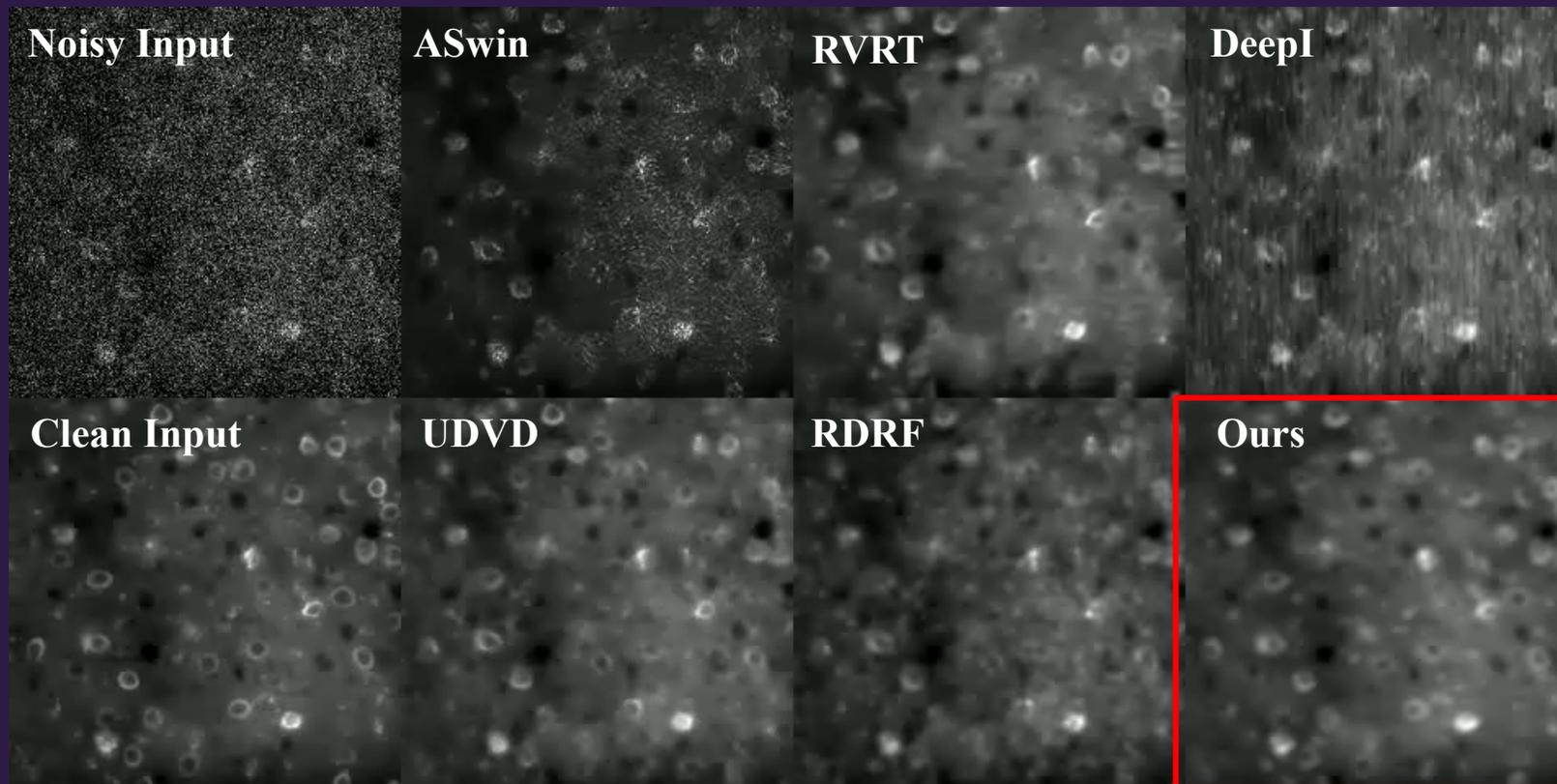
[4] Song et al., Neural anatomy and optical microscopy (naomi) simulation for evaluating calcium imaging methods. Journal of neuroscience methods 2

[5] Liang et al., Recurrent video restoration transformer with guided deformable attention. Neurips 2022

[6] Lindner et al., Lightweight video denoising using aggregated shifted window attention. ICCV, 2023.

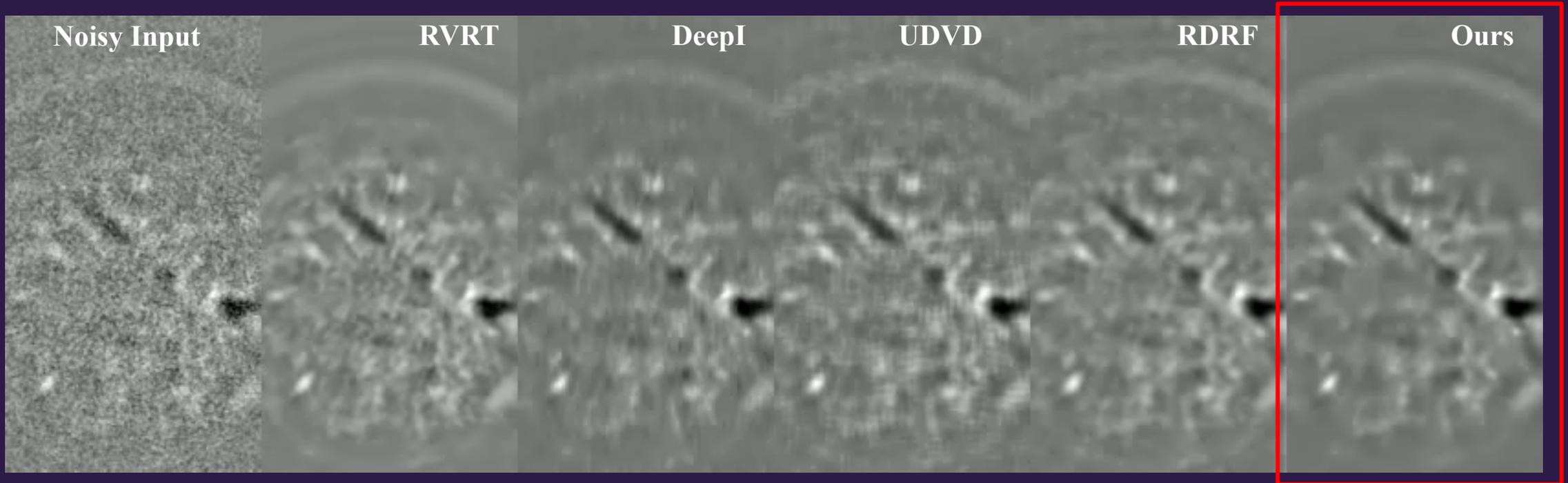
Results

Simulated two-photon calcium imaging



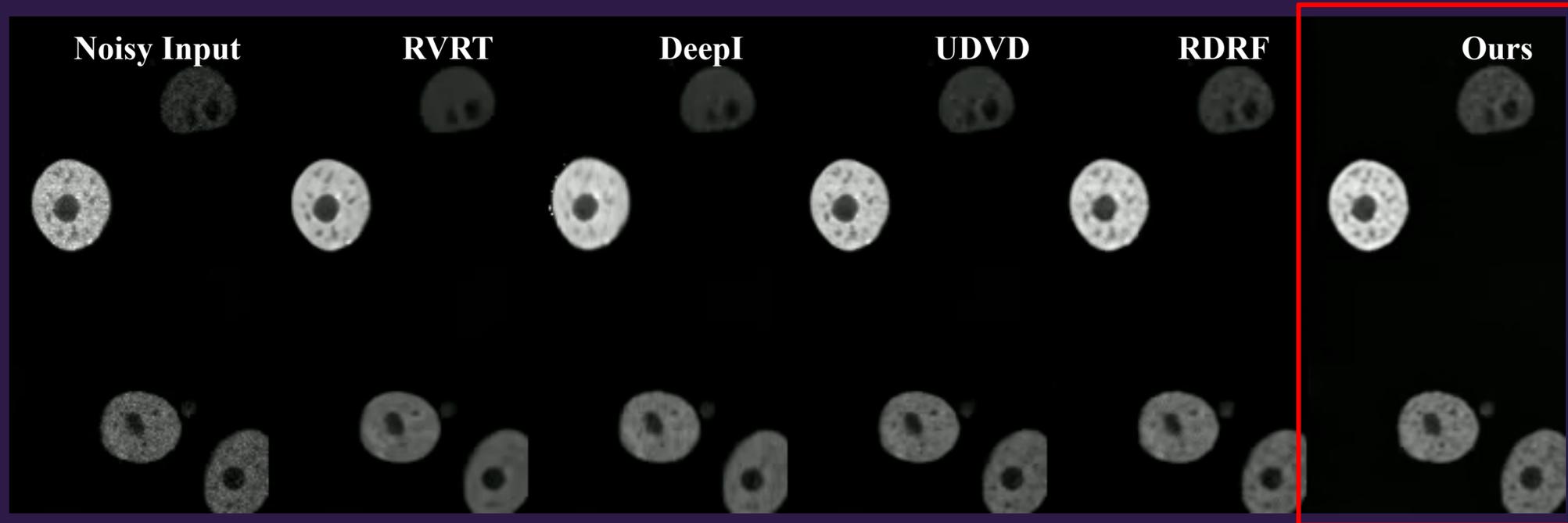
Results

One-photon calcium imaging



Results

Fluorescence Microscopy

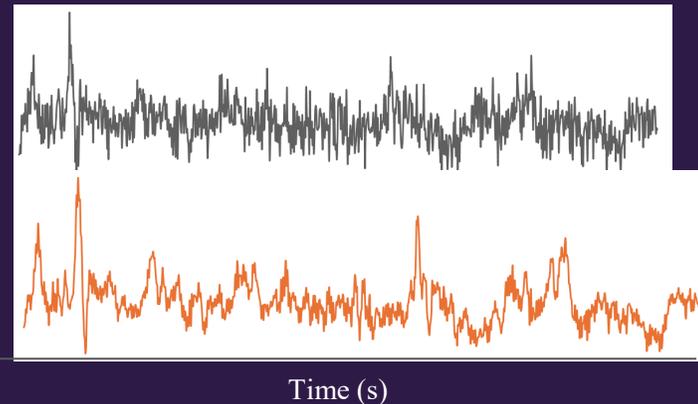
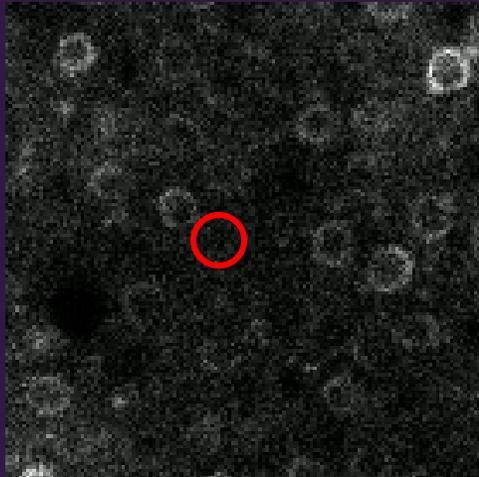


Results

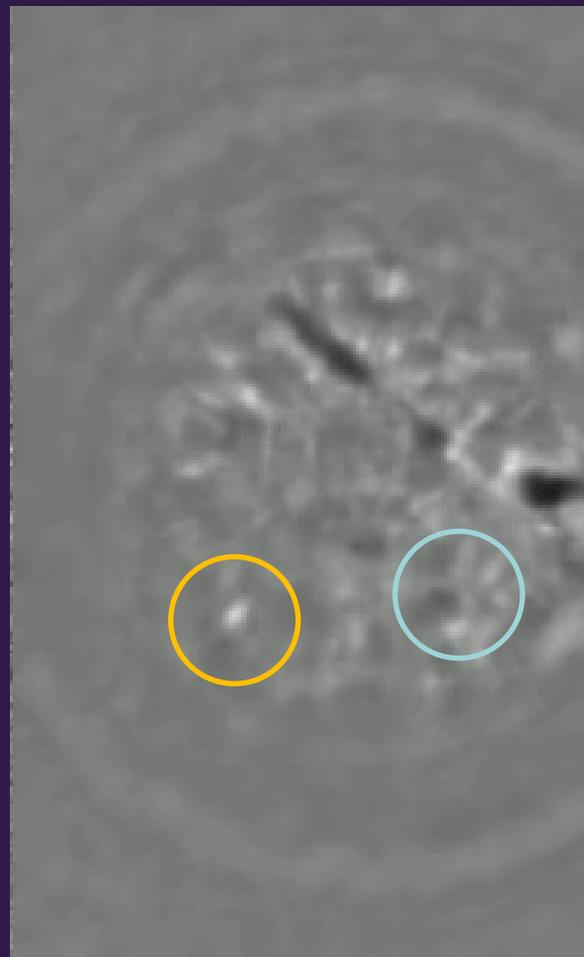
- The result shows the average PSNR and SSIM values on the 2P imaging simulated with NAOMi¹

	RVRT	ASwin	DeepI	UDVD	RDRF	Ours
PSNR	<u>30.05</u>	25.94	28.64	25.75	28.31	35.90
SSIM	<u>0.92</u>	0.90	0.91	0.74	<u>0.92</u>	0.95

- Calcium traces of an example ROI from the two-photon calcium imaging

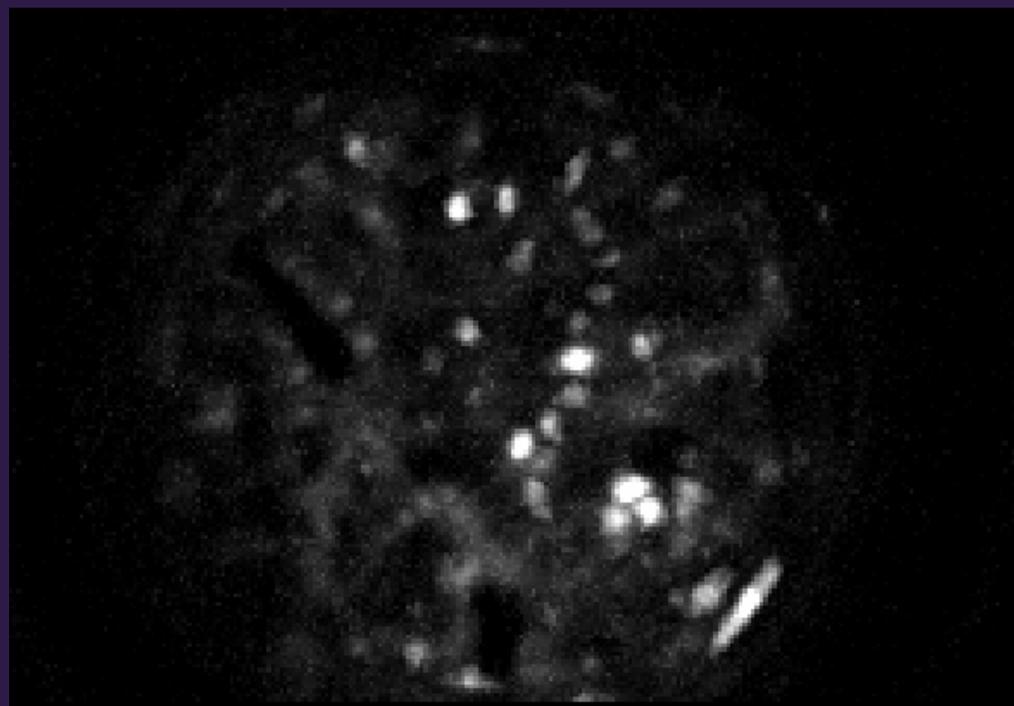


Results

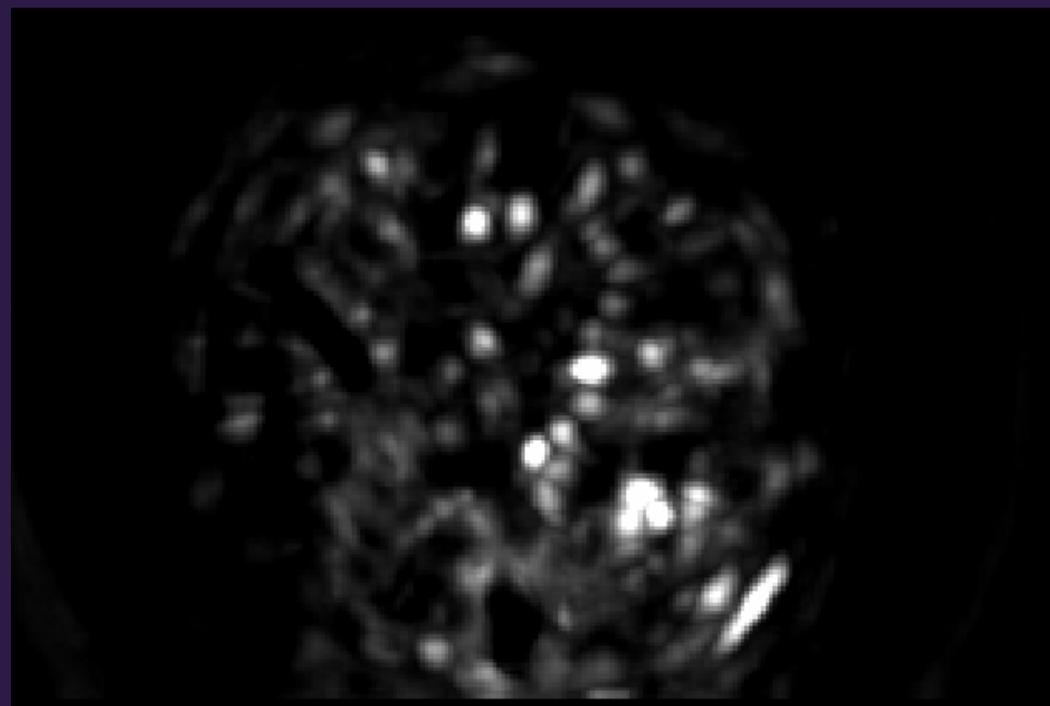


Effectively
removed the
noise while
preserved the
subtle neuron
activity!

Maximum Projection



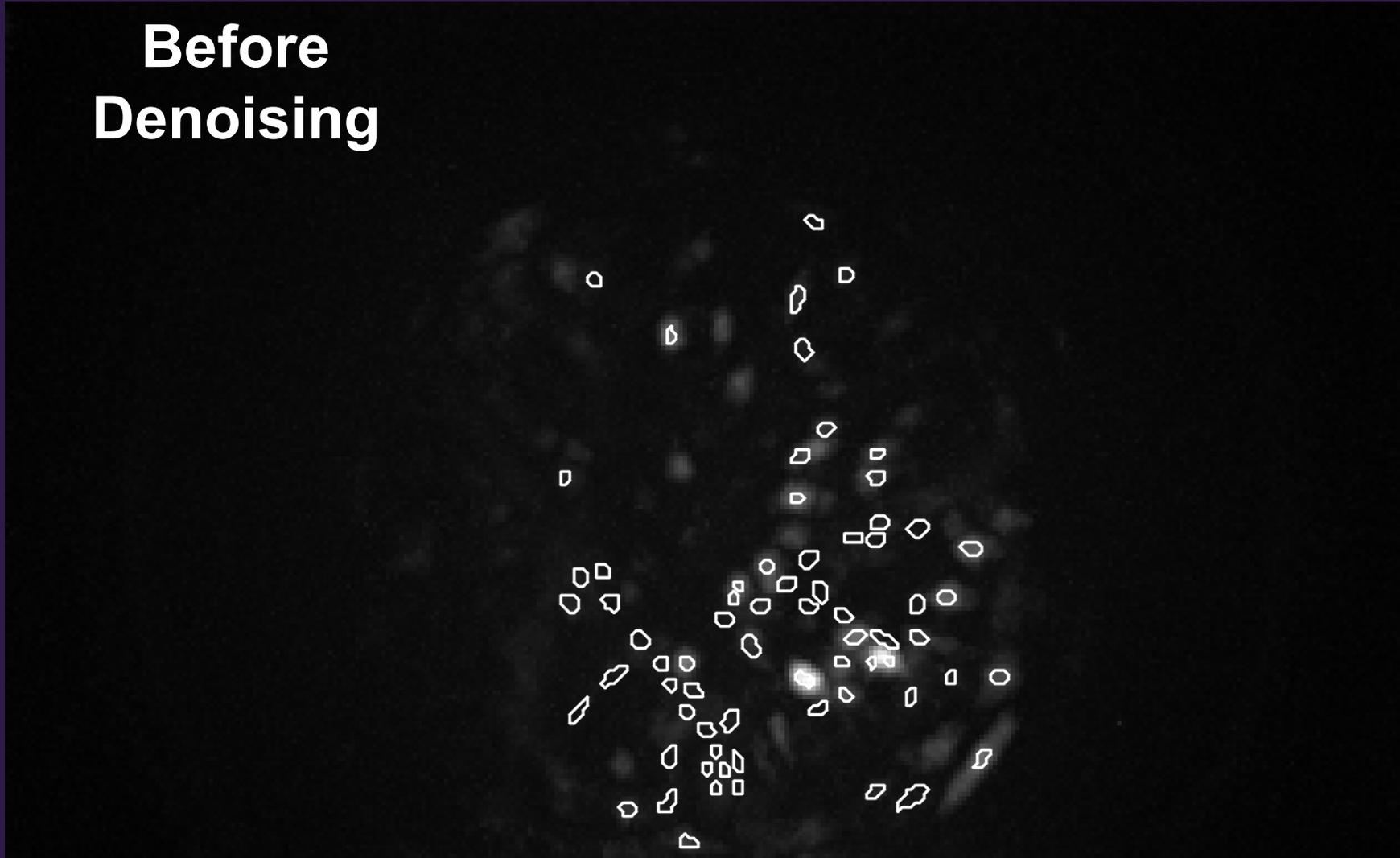
Original



Ours

Results – Cell Tracking Rate

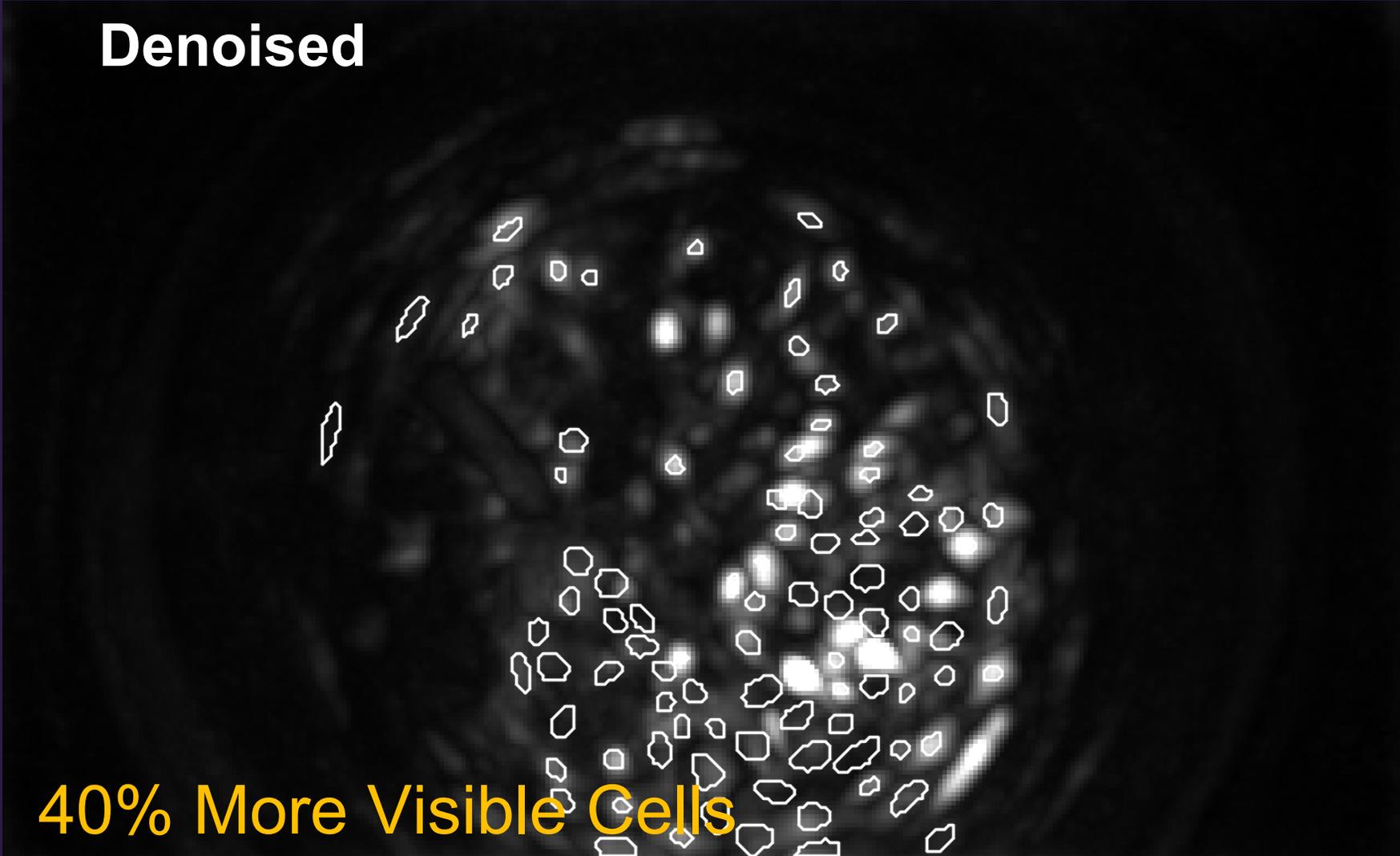
**Before
Denoising**



Results – Cell Tracking Rate

Denoised

40% More Visible Cells

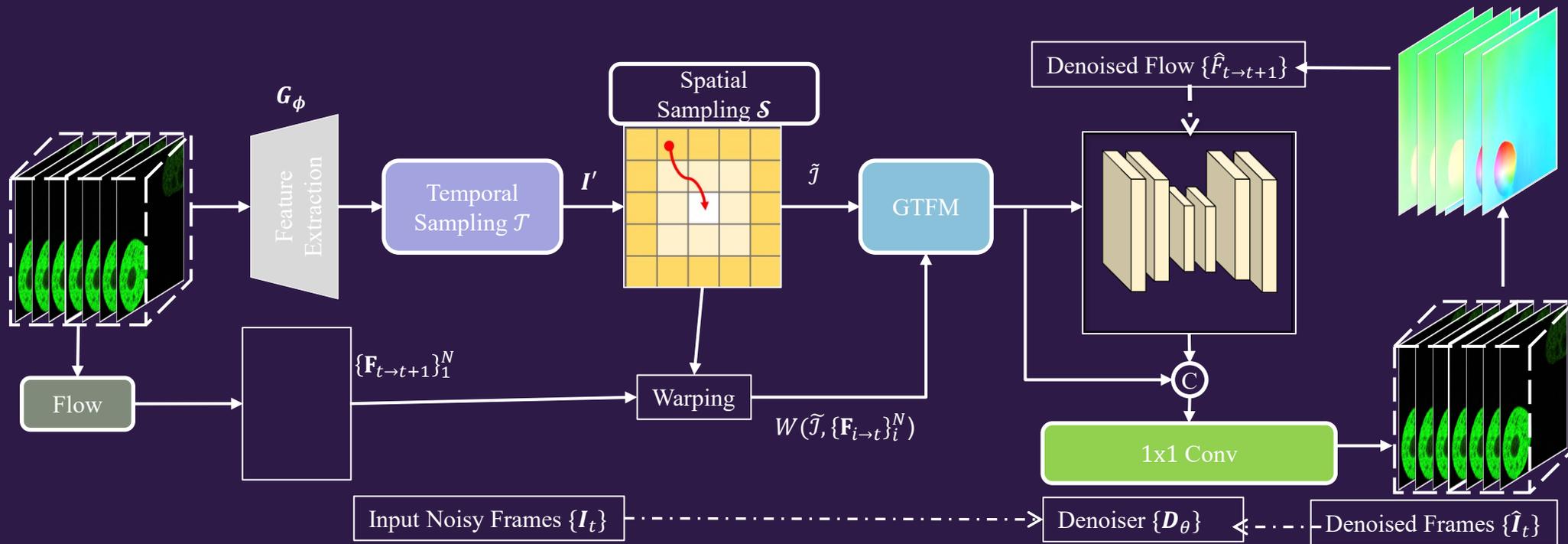
A grayscale microscopy image showing a dense population of cells. The cells are represented by white outlines, indicating they have been successfully tracked. The background is dark with some faint, blurry structures. The text 'Denoised' is in the top left, and '40% More Visible Cells' is in the bottom left.

Conclusion

- We presented **a novel unsupervised denoising method** utilizing DeepTemporal Interpolation.
- Our approach demonstrates **strong generalization** across a variety of **medical imaging** and color datasets and shows robustness to different noise types and intensities.
- **Limiting motion handling**, due to the fixed temporal filter.

Unsupervised Denoising for Multi-Modal Microscopy Imaging for Small Data Scenarios

Method



Method

Weighted Temporal Sampling

1. Given the Magnitude \bar{M} of optical flow, $\{F_{i \rightarrow t}\}_i^N$, we compute the initial minimum weight κ_0 .

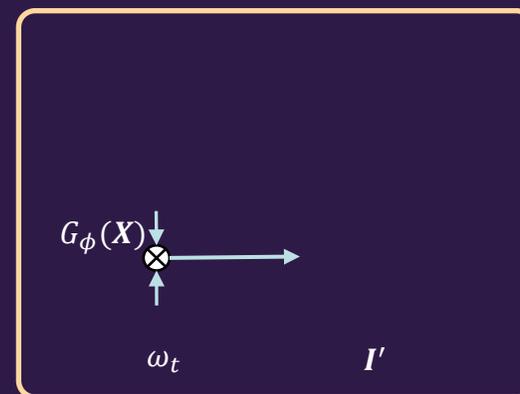
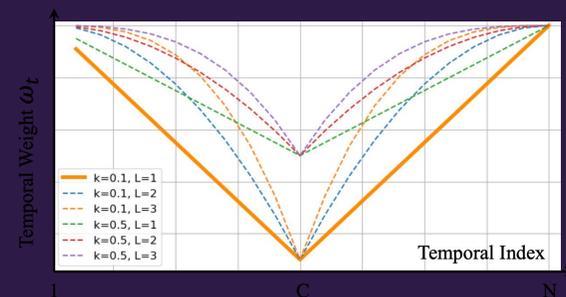
$$\kappa_0 = 0.2 \cdot (1 - e^{-\eta \times \bar{M}}) \quad \kappa = \max\left(0, \kappa_0 \left(1 - \frac{\text{epoch}}{\text{max_epoch}}\right)\right)$$

2. We compute the temporal weights $\{\omega_t | t = 1, \dots, N\}$

$$\omega_t = \begin{cases} \kappa + (1 - \kappa) \left(1 - \left(\frac{t}{C}\right)^L\right), & \text{if } t \leq C \\ \kappa + (1 - \kappa) \left(1 - \left(\frac{2C - t}{C}\right)^L\right), & \text{otherwise,} \end{cases}$$

3. We apply the temporal weights to the features

$$I'_t = \omega_t \odot G_\phi(X_t) \quad \forall t \in 1, \dots, N$$

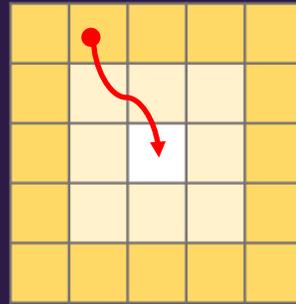


Method

Weighted Spatial Sampling

- Replaces randomly selected pixels p with neighboring pixels q

$$P(q|p) = \frac{e^{-\alpha \cdot d(p,q)}}{\sum_{q' \in \Omega_p} e^{-\alpha \cdot d(p,q')}}$$



where:

$P(q|p)$ is the probability of choosing a neighboring pixel

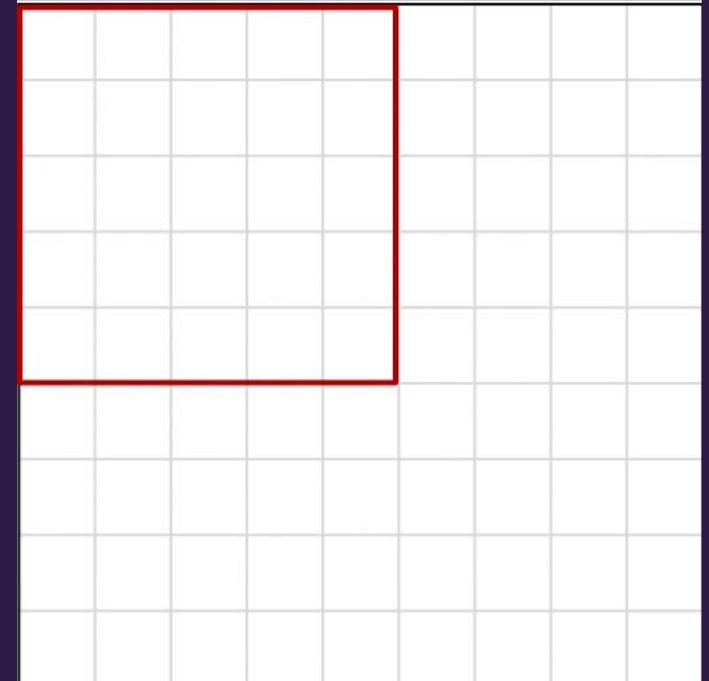
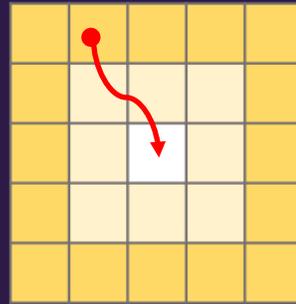
$d(p, q)$ is the l_1 distance between pixels p and q

Method

Weighted Spatial Sampling

- Replaces randomly selected pixels p with neighboring pixels q

$$P(q|p) = \frac{e^{-\alpha \cdot d(p,q)}}{\sum_{q' \in \Omega_p} e^{-\alpha \cdot d(p,q')}}$$



where:

$P(q|p)$ is the probability of choosing a neighboring pixel

$d(p, q)$ is the l_1 distance between pixels \mathbf{p} and \mathbf{q}

Method

Guided Temporal Fusion

1. Given spatiotemporal sampled features, we first warp the neighboring features to the central frame and the mean warped \bar{W} .

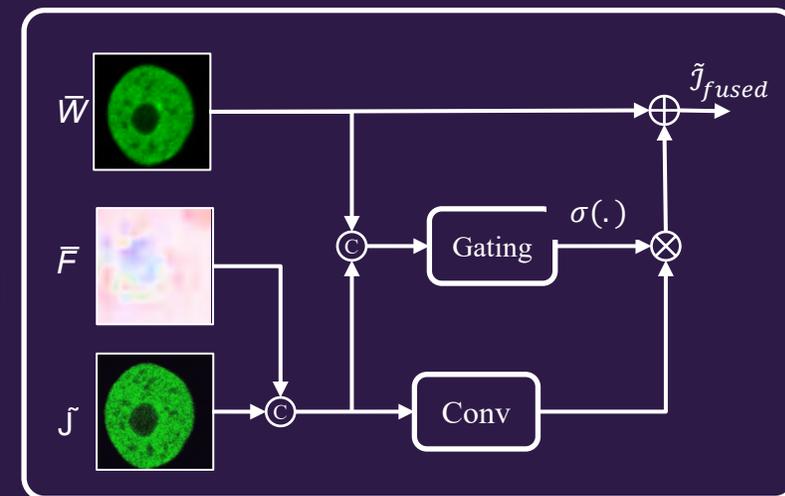
$$\bar{W} = \frac{1}{N-1} \sum_{i \neq t}^N \text{warp}(\tilde{J}_i, \mathbf{F}_{i \rightarrow t})$$

2. We concatenate the mean flow \bar{F} , mean warped \bar{W} and the central feature and pass to gating network to yield spatial attention map $\sigma(\cdot)$

3. We concatenate the mean flow \bar{F} and the central feature through convolution to get residual candidate.

4. Finally, we fused the temporal features using:

$$\tilde{J}_{fused} = \bar{W} + \sigma * \Delta \tilde{J}_t$$



Method

Recurrent Optimization

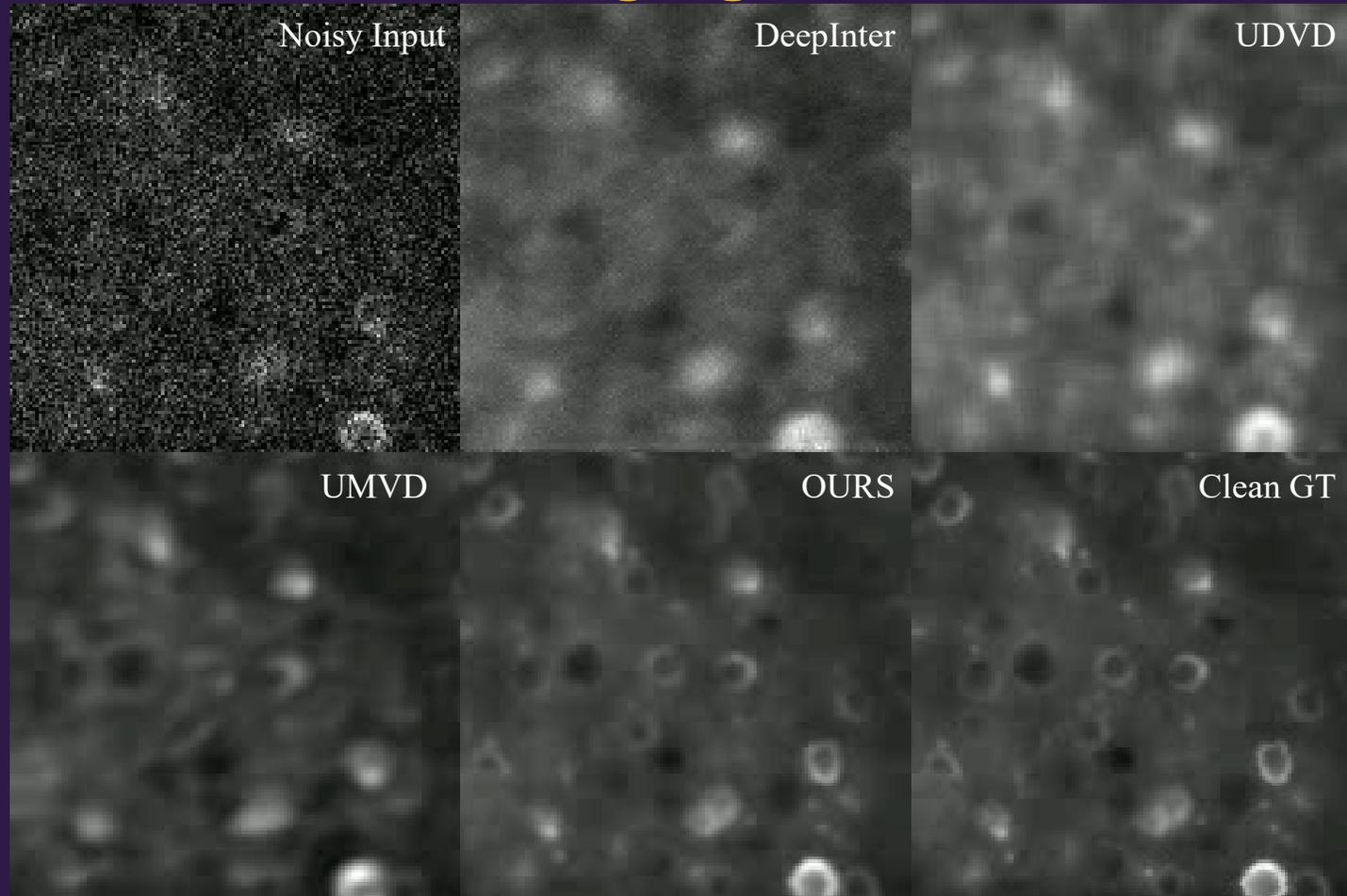
- ❖ To effectively guide the network during training, we employ two loss terms. One is the pixel-wise L1 loss between \hat{I}_t and the reference frame I_t^{ref} , and the second loss term is a consistency loss based on the optical flow estimation of the denoised frames.

$$\mathcal{L}_1 = \sum_{p \in I_t} |\hat{I}_t(p) - I_t^{ref}(p)| \quad \mathcal{L}_2 = \frac{1}{N} \sum_{t=1}^{N-1} \|W(\hat{I}_{t+1}, \hat{F}_{t \rightarrow t+1}) - \hat{I}_t\|^2$$

$$\mathcal{L} = \mathcal{L}_1 + \mathcal{L}_2$$

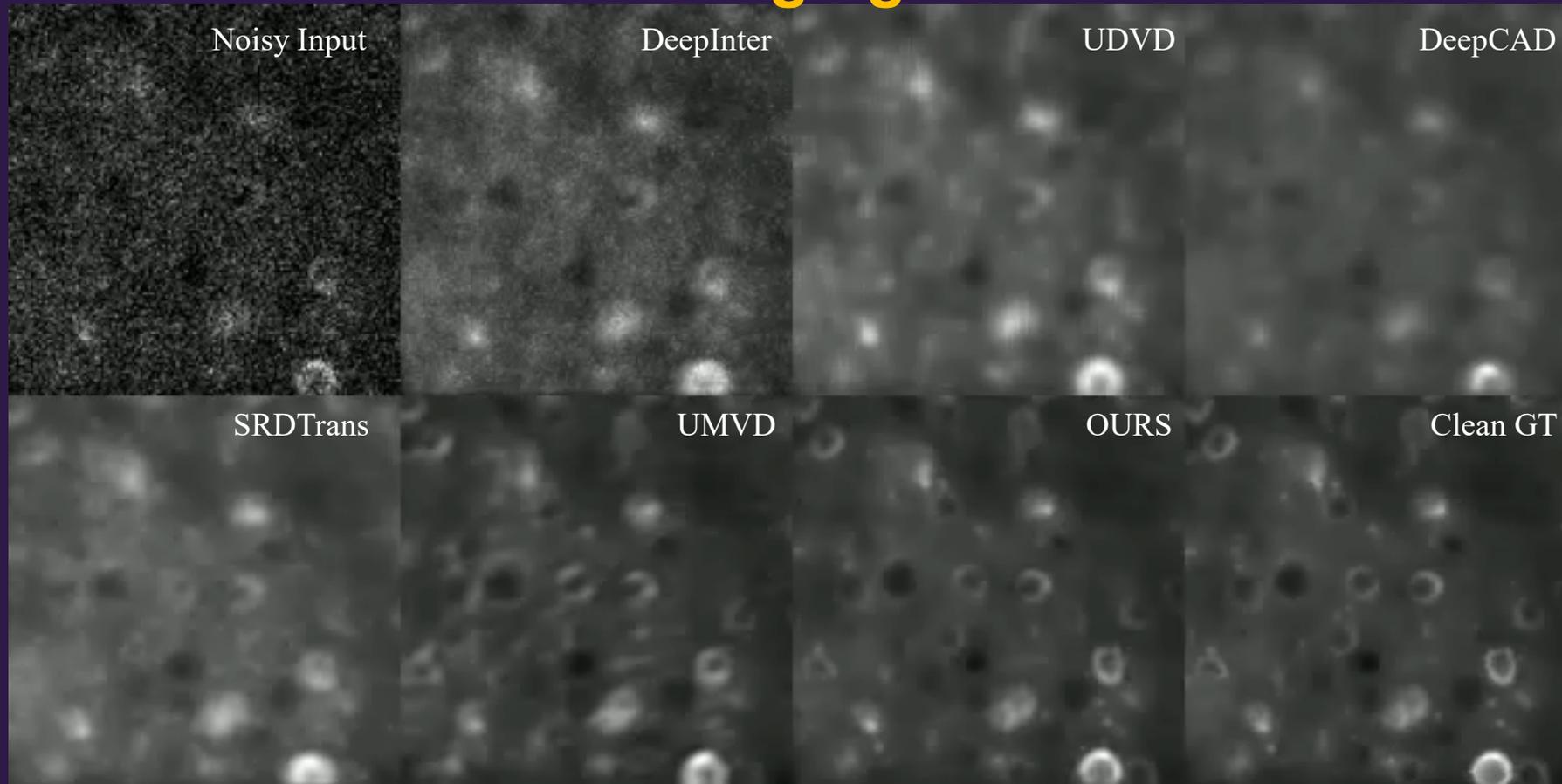
Result

Calcium Imaging – 10 Frames



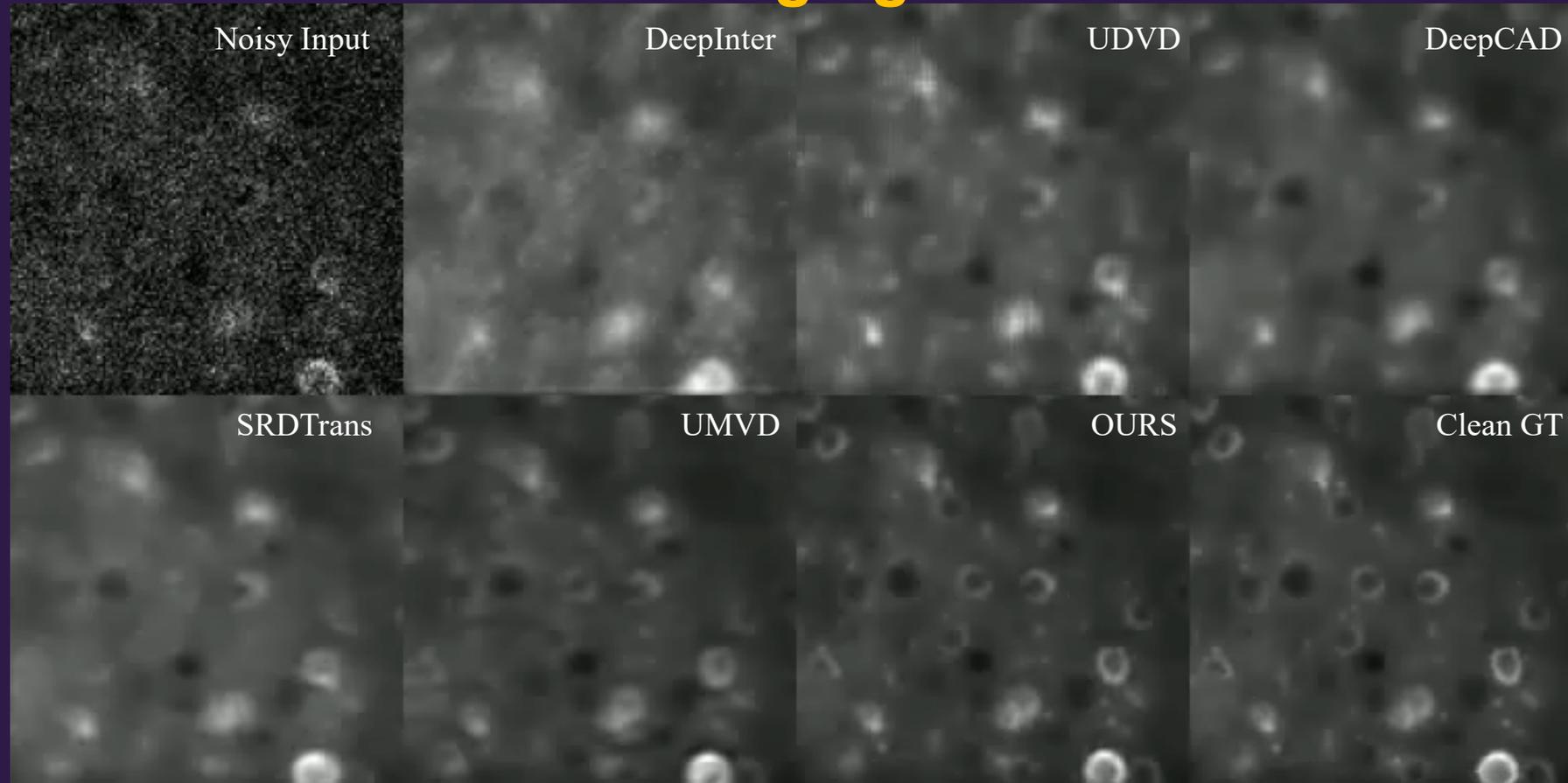
Result

Calcium Imaging – 20 Frames



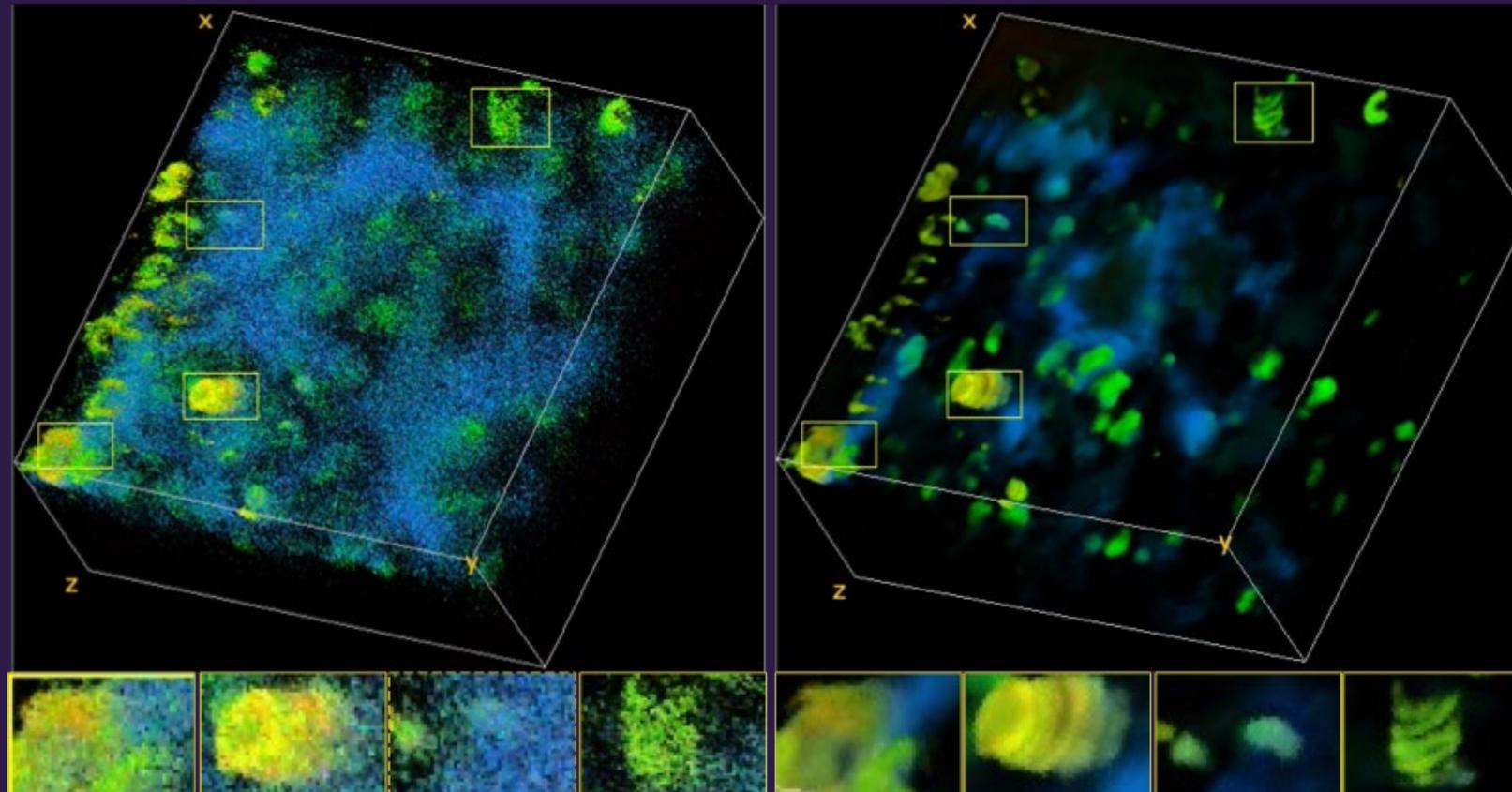
Result

Calcium Imaging – 50 Frames



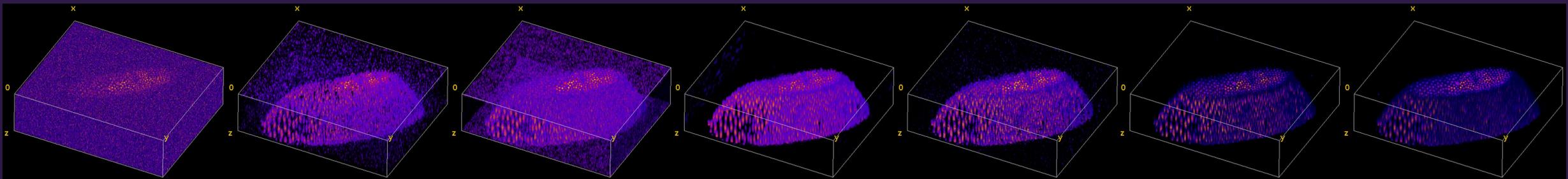
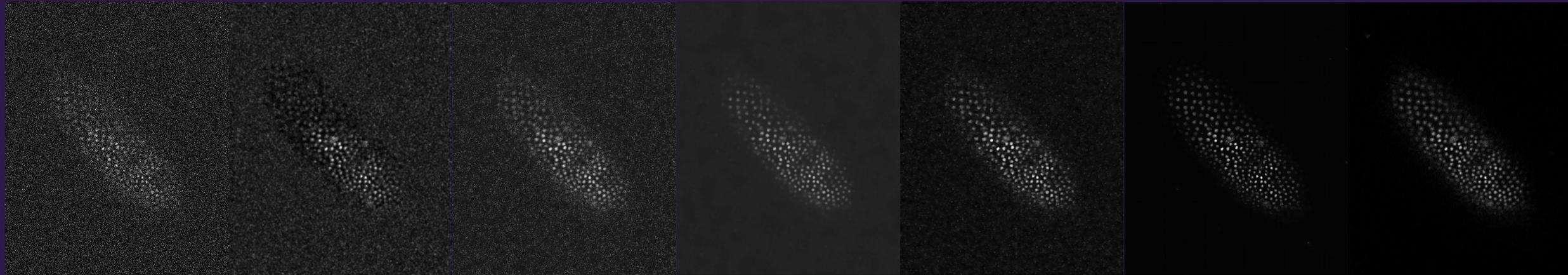
Result

Confocal Imaging



Results

Confocal Imaging



Noisy Input

DeepCad

DeepInterpolation

UDVD

UMVD

SRDTrans

Ours

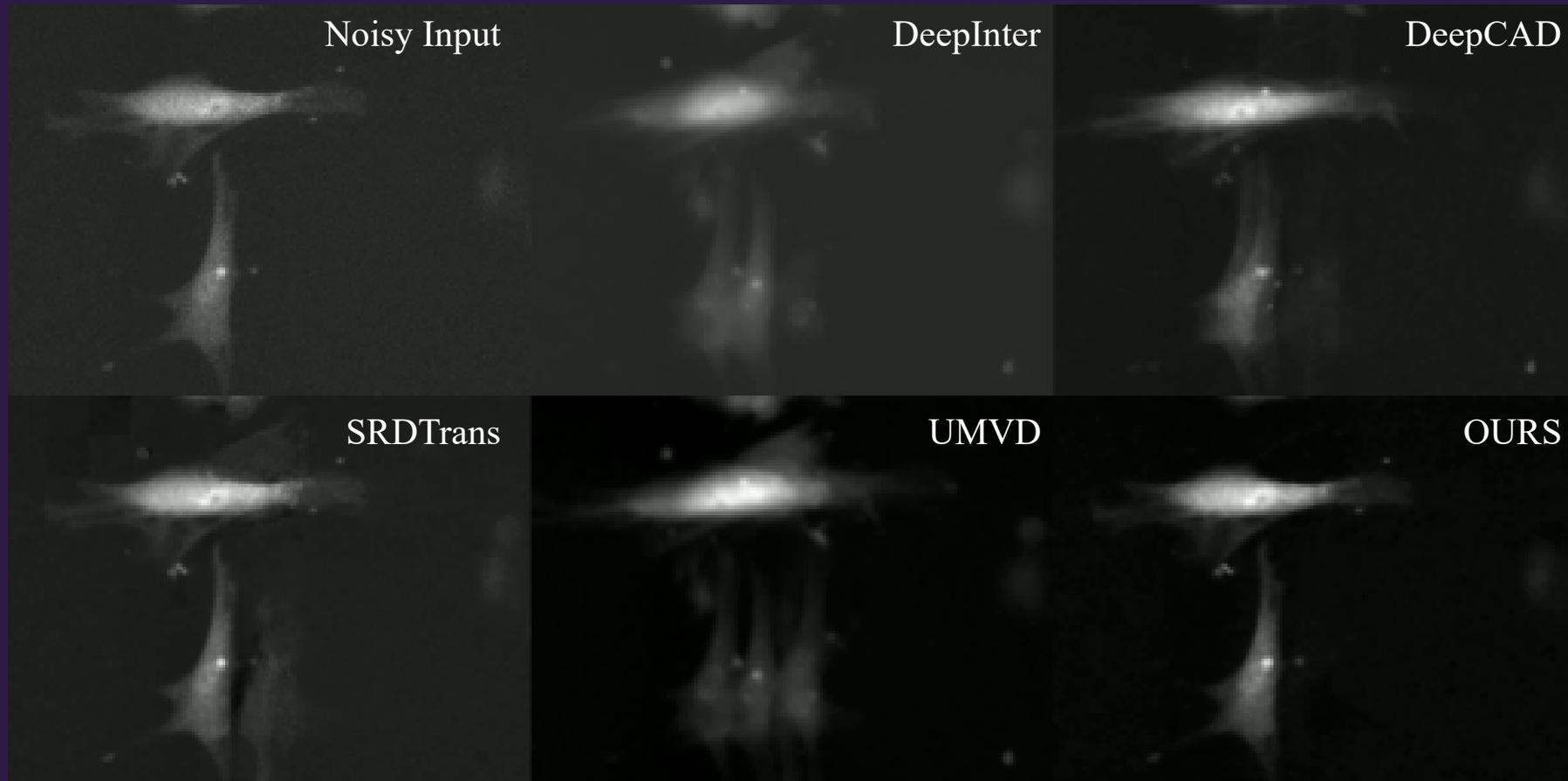
Result

Fluorescence Microscopy



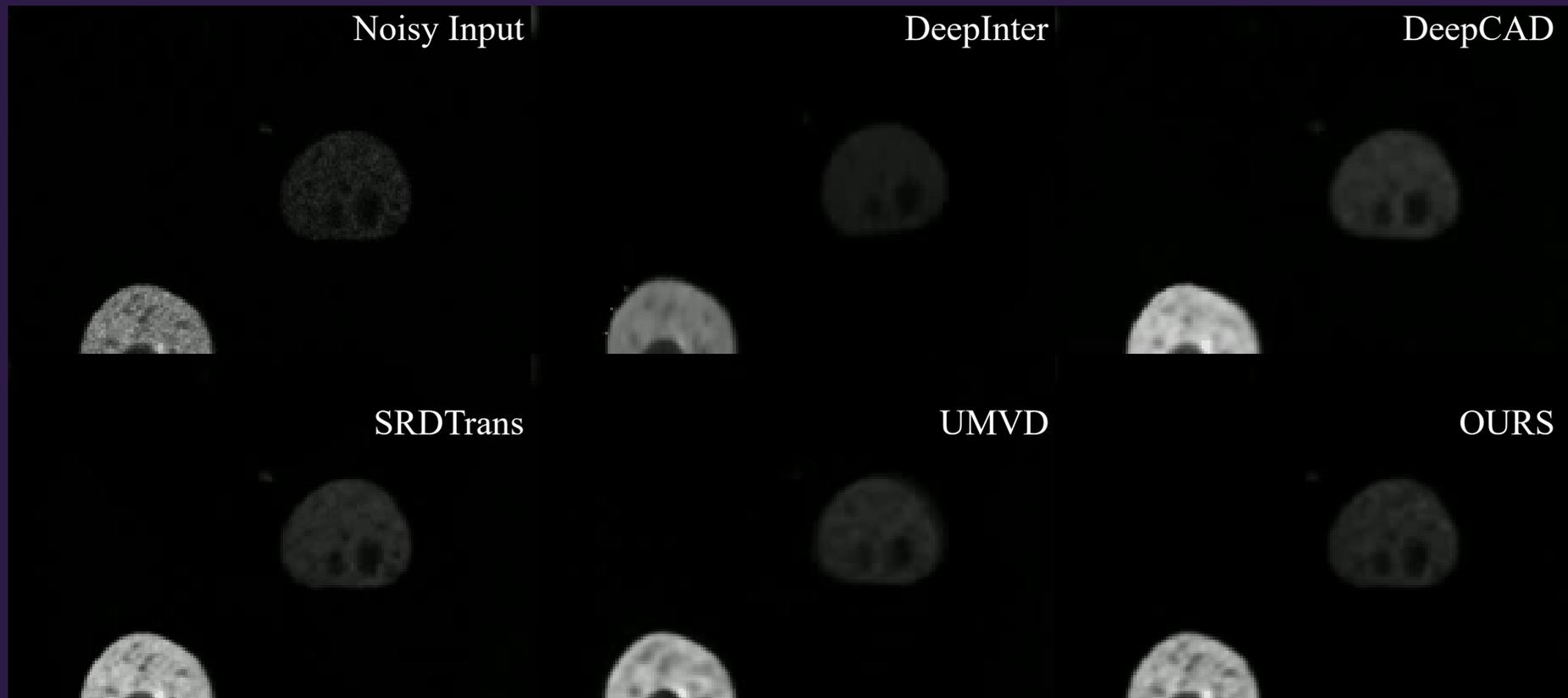
Result

Fluorescence Microscopy



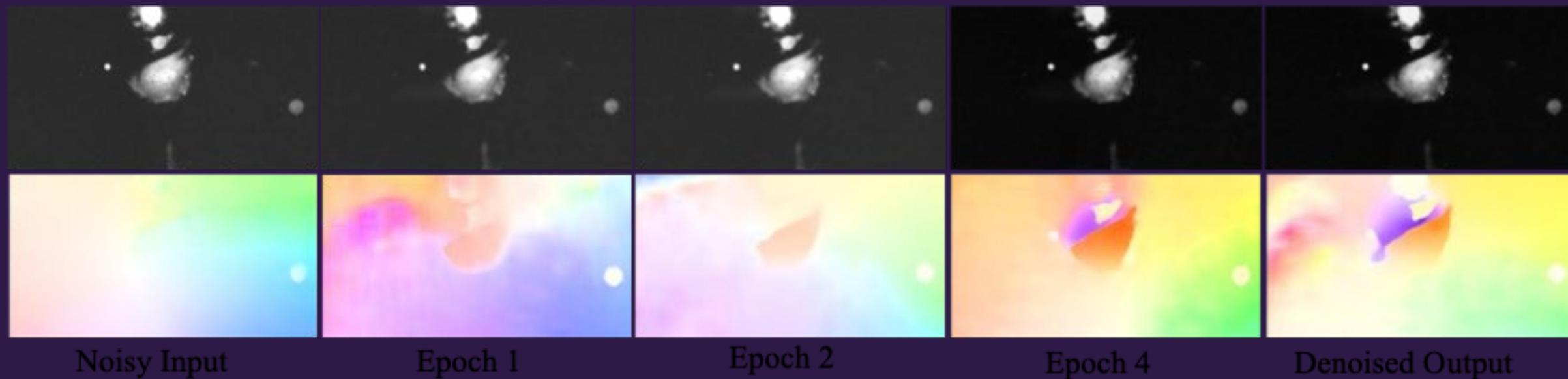
Result

Fluorescence Microscopy



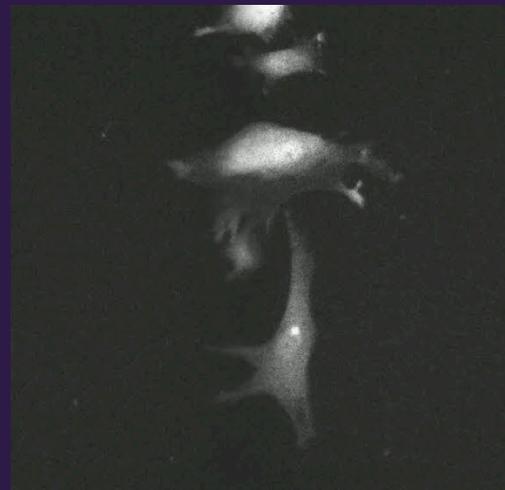
Result

Refined Flow



Result

Optical Flow



Noisy Input



Denoised



Noisy Flow



Denoised Flow

Quantitative Results

- ❖ Quantitative comparison with SOTA video denoising techniques on simulated two-photon calcium imaging.

	DeepI		DeepCAD		UDVD		SRDTrans		UMVD		Ours	
	PSNR \uparrow	SSIM \uparrow										
150 μm^2	30.020	0.911	36.606	0.971	31.889	0.899	41.176	0.984	38.871	0.985	43.970	0.992
500 μm^2	34.222	0.907	29.130	0.885	35.775	0.932	34.595	0.924	32.933	0.909	37.501	0.952
Average	32.121	0.909	32.868	0.928	33.832	0.916	<u>37.886</u>	<u>0.954</u>	35.902	0.947	40.736	0.972

Conclusion

- ❖ We presented a novel **unsupervised** denoising method which dynamically adapts to observed motion in videos by utilizing weighted SpatioTemporal sampling.
- ❖ Our approach demonstrates strong generalization across a variety of medical imaging datasets.
- ❖ To improve scalability to high-resolution or real-time settings, future work will explore lightweight optical flow models and adaptive sampling strategies.

Thanks to all Collaborators and Sponsors!



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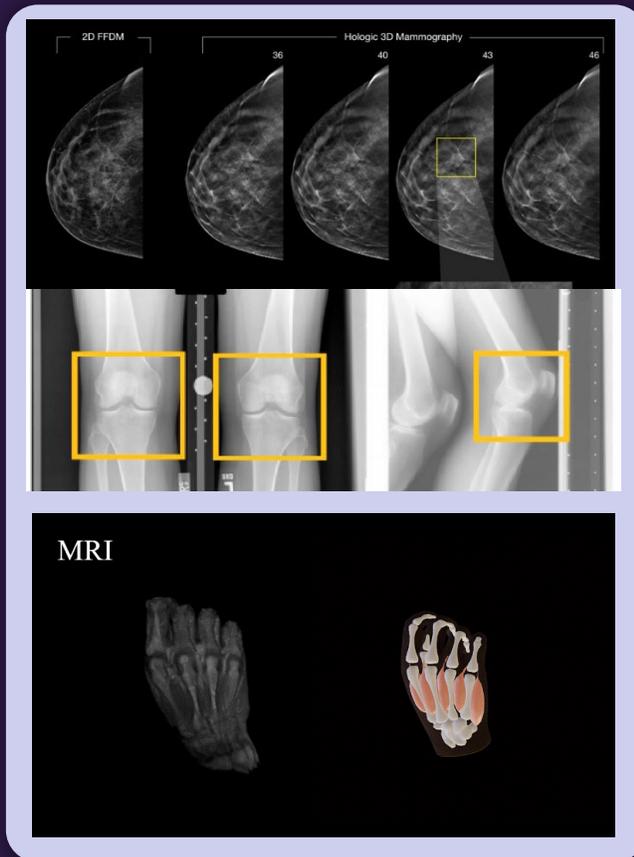


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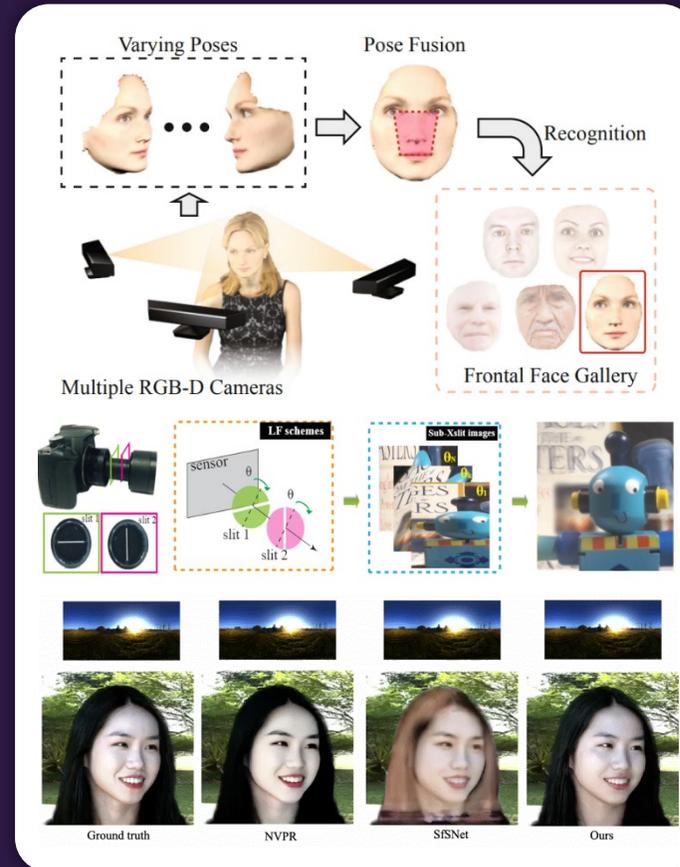


Advanced Imaging Laboratory

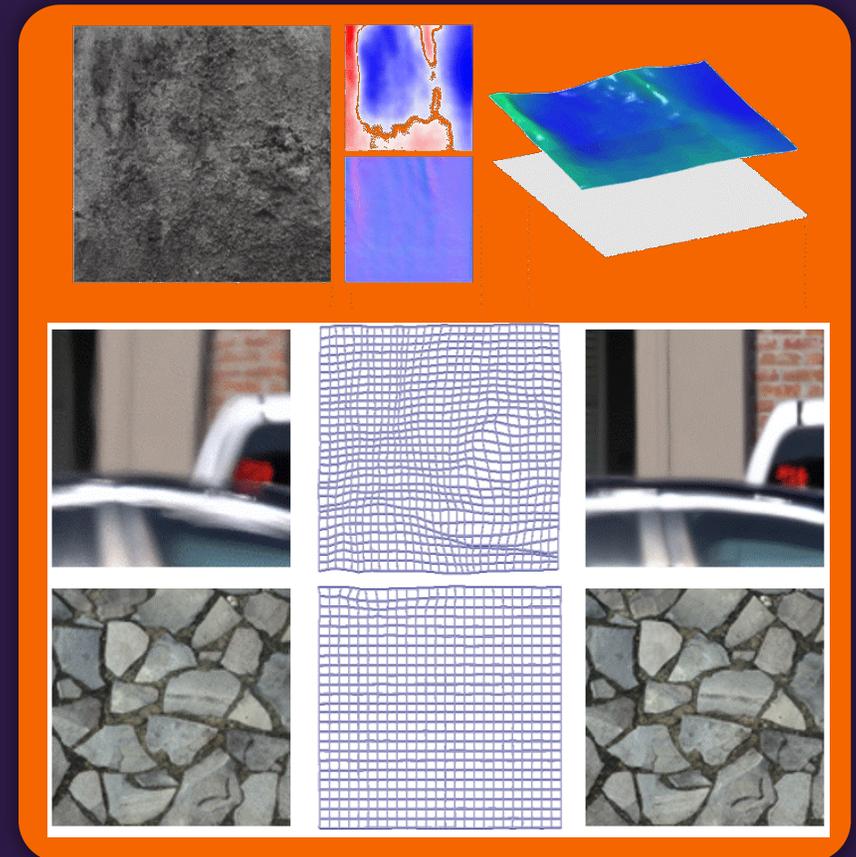
Medical Image Processing



Computer Vision



Physics-informed Machine Learning



THANK YOU

Project Website



<https://nianyil.people.clemson.edu/>